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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶:
A01N 63/00, 43/04, C12N 15/00, C07H 21/02

(11) International Publication Number:

WO 99/26480

A1

(43) International Publication Date:

3 June 1999 (03.06.99)

(21) International Application Number:

PCT/US98/24950

(22) International Filing Date:

20 November 1998 (20.11.98)

(30) Priority Data:

08/975,424

20 November 1997 (20.11.97) US

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(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published

With international search report.

(54) Title: ANTI-ANGIOGENIC GENE THERAPY VECTORS AND THEIR USE IN TREATING ANGIOGENESIS-RELATED DISEASES

(57) Abstract

A method for inhibiting tumor growth in a human patient harboring a solid tumor, said method comprising administering to said patient a nucleic acid molecule which expresses in said patient an anti-angiogenic polypeptide selected from the group consisting of human angiostatin, murine angiostatin, human endostatin, murine endostatin, and angiogenesis-inhibiting fragments thereof, wherein expression of the anti-angiogenic polypeptide in the patient inhibits angiogenesis in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells, thereby inhibiting its growth.

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ANTI-ANGIOGENIC GENE THERAPY VECTORS AND THEIR USE IN TREATING ANGIOGENESIS-RELATED DISEASES

Field of the Invention

This invention relates generally to gene therapy for, e.g., cancer.

Background of the Invention

Angiogenesis is the process by which new capillaries are formed from existing vasculature. It is a complex process which involves proliferation and migration of endothelial cells. It plays a fundamental role in reproduction, development and wound repair. Unregulated angiogenesis, however, can further the progression of many diseases, including tumor growth and metastasis, arthritis, diabetes, and some forms of blindness. For example, there is experimental evidence that limits of tumor size and growth are not the failure of the tumor cells to proliferate, but rather a failure of the tumor to provide sufficient nutrients and waste removal to its constituent cells by recruiting surrounding vasculature.

Summary of the Invention

The invention features a method for inhibiting tumor growth in a

human patient harboring a solid tumor, involving administering to the patient a
nucleic acid molecule which expresses in the patient an anti-angiogenic
polypeptide selected from the group consisting of human angiostatin, murine
angiostatin, human endostatin, murine endostatin, and angiogenesis-inhibiting
fragments thereof, wherein expression of the anti-angiogenic polypeptide in the

patient inhibits angiogenesis in the vicinity of the tumor and/or systemically by
diffusion of the recombinant protein to the vascular compartment from
secreting transduced cells, thereby inhibiting its growth.

In a second, related aspect, the invention features tumor inhibition, of

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the type just described, using nucleic acids molecules of the formula A-B, where A and B are polypeptide and/or export signal joined by a peptide bond; peptide A contains at least 100 amino acids and includes at least kringles 1, 2, and 3 of human or murine angiostatin; and peptide B contains at least 100 amino acids and includes at least 75% of the amino acid sequence of human or murine endostatin. Expression of the fusion anti-angiogenic polypeptide in the patient inhibits angiogenesis in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells, thereby inhibiting its growth. In some embodiments of this hybrid polypeptide and/or export signal method, polypeptide and/or export signal A further includes kringle region 4 of angiostatin, and can also include kringle region 5 of plasminogen (the larger protein molecule of which angiostatin is a portion).

In both aspects of the invention, the nucleic acid molecule preferably constitutes a portion of a viral vector or a plasmid, which can either be administered to the patient so that cells of the patient in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells are infected or transfected with the nucleic acid encoding the angiogenesis-inhibiting polypeptide, or cells (of the patient, or another human donor, or an animal) are infected or transfected *ex-vivo*, and those infected or transfected cells are then infused into the patient so that the anti-angiogenic polypeptide is expressed in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells.

As will be discussed in more detail below, in particularly effective embodiments, the nucleic acid molecule includes a nucleotide sequence

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encoding a preactivation polypeptide and/or export signal for effecting Golgi and/or endoplasmic reticulum export of the anti-angiogenic polypeptide.

In another aspect, the invention features a method for treating a human patient suffering from diabetic retinopathy, involving administering to the patient one of the nucleic acid molecules described above.

The above and other features, objects and advantages of the present invention will be better understood by a reading of the following specification in conjunction with the drawings.

Brief Description of the Drawings

Fig. 1 depicts the structural relationship of angiostatin with plasminogen.

Fig. 2 depicts the structural relationship of endostatin with collagen type XVIII.

Fig. 3 depicts various viral (A. MSCV murine retrovirus; B. Adenoassociated virus; C. HIV based retrovirus; E. recombinant adeno-virus) and
non-viral (D. plasmid) vectors used in the construction of gene therapy vectors
for this invention.

-4-

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47

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MSCV: Murine Stem Cell Virus LTR: Long Terminal Repeat Rous Sarcoma Virus RSV: Inverted Terminal Repeat ITR: Human Immunodeficiency Virus HIV: Internal Ribosomal Entry Site IRES: Green Fluorescence Protein GFP: HBPRE: Hepatitis B Export Element Rev Response Element RRE: polyA: polyadenylation site , viral packaging sequence

The inverted triangle shows the site at which the anti-angiogenic constructs will be inserted using engineered MluI and XhoI restriction sites.

The arrow denotes the direction of transcription.

Fig. 4 depicts in the left (A) panel nude mice which were implanted with human neuroblastoma cells (line SK-N-AS) transduced with a mock virus and in the right (B) panel, nude mice which were transplanted with human neuroblastoma cells transduced with a retroviral gene therapy vector encoding an angiostatin-endostatin fusion protein.

Fig. 5 shows the nucleotide sequence (SEQ ID NO: 1) and amino

^{*} denotes specific mutations within the long terminal repeat and leader which bestows the ability for expression in embryonic stem and hematopoietic stem cells.

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acid sequence (SEQ ID NO: 2) of human plasminogen and the nucleotide sequence (SEQ ID NO: 5) and amino acid sequence (SEQ ID NO: 6) of human angiostatin.

Fig. 6 shows the nucleotide sequence (SEQ ID NO: 9) and amino acid sequence (SEQ ID NO: 10) of murine endostatin.

Fig. 7 shows the nucleotide sequence (SEQ ID NO: 3) and amino acid sequence (SEQ ID NO: 4) of murine plasminogen and the nucleotide (SEQ ID NO: 7) and amino acid sequence (SEQ ID NO: 8) of murine angiostatin.

Detailed Description

This invention provides gene therapy using a vector having a nucleotide sequence encoding one of the above-identified anti-angiogenic polypeptides. Described below in more detail are some of the components of the vectors and methods of the invention.

By a gene therapy vector is meant a vector useful for gene therapy. Gene therapy vectors carry a gene of interest that is useful for gene therapy. The gene therapy vectors are able to be transferred to the cells of an animal, e.g., a human, and are able to express the gene of interest in such cells so as to effect gene therapy. The vector can be, e.g., chromosomal, non-chromosomal, or synthetic, and can be RNA or DNA. The vector can be, e.g., a plasmid, a virus or a phage. Preferred vectors include, e.g., retroviral vectors, adenoviral vectors, adeno-associated vectors, herpes virus vectors, Simliki Forest Virus-based vector, Human Immunodeficiency virus, Simian Immunodeficiency virus, and non-viral plasmids. A preferred retroviral vector is Murine Stem Cell Virus (MSCV), which is a variant of Moloney Murine Leukemia Virus (MoMLV).

By anti-angiogenic polypeptide is meant a polypeptide which inhibits

-6-

angiogenesis. The terms polypeptide, protein and polypeptide and/or export signal are used interchangeably herein. By angiogenesis is meant the process by which new vasculature, in particular, new capillaries, are formed from existing vasculature. Angiogenesis is a complex process entailing numerous steps, including local dissolution of the basement membrane, migration of endothelial cells into the surrounding stroma, proliferation of the endothelial cells at the leading edge to form a migrating column of cells, branching and fusion of the newly formed vascular loops, and formation of a new basement membrane. By inhibiting angiogenesis is meant completely or partially inhibiting the formation of such new vasculature.

In certain embodiments, the anti-angiogenic polypeptide is an antiangiogenic fragment of plasminogen (in particular, angiostatin), an antiangiogenic fragment of collagen XVIII (endostatin) or a fusion of the two fragments.

Angiostatin is an internal fragment of plasminogen having a molecular weight of 38 or 45 kDa, depending on whether it contains kringles 1-3 or 1-4. In the invention, either can be used, or a molecule including kringles 1-3 and a portion of kringle 4 can be used. Angiostatin can be naturally produced in vivo in small amounts by tumor cells, e.g. murine Lewis lung carcinoma cells, by proteolytic cleavage of plasminogen so as to eliminate the N-terminal portion including the signal polypeptide and/or export signal and the preactivation polypeptide and/or export signal, as well as the C-terminal portion following kringle 3 or 4. Mouse and human angiostatin have been purified and sequenced. In preferred embodiments, the gene therapy vectors of this invention encode angiostatin having kringles 1, 2 and 3, or angiostatin having kringles 1, 2, 3 and 4.

In another preferred embodiment, the anti-angiogenic polypeptide is

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endostatin or a biologically active analog or fragment thereof. Endostatin can be naturally produced *in vivo* in small amounts by tumor cells, e.g., murine angiosarcoma cells, by proteolytic cleavage of endogenous collagen XVIII so as to eliminate the N-terminal portion including the signal polypeptide and/or export signal and the preactivation polypeptide and/or export signal, as well as the C-terminal portion following kringle 3 or 4. See Fig.2. Mouse endostatin has been sequenced, and the human molecule (SEQ ID NOs: 17 and 18) forms a portion of collagen 18 (SEQ ID NOs: 19 and 20).

The human molecule position and sequence are apparent from an alignment of the active, Lys-terminated active region of human collagen 18 with murine endostatin, such that the C-terminal lysine residues align, bringing the active endostatin sequences into alignment.

In yet another preferred embodiment, the anti-angiogenic polypeptide is an in-frame fusion of angiostatin or a biologically active analog or fragment thereof and endostatin or a biologically active analog or fragment thereof. Preferably, the angiostatin or biologically active analog or fragment is 5' of the endostatin or biologically active analog or fragment. In certain embodiments, the angiostatin-endostatin fusion proteins exhibit synergistic anti-angiogenic properties.

By fragment is meant some portion of the naturally occurring antiangiogenic polypeptide. Preferably, the fragment is at least 20 amino acid residues, more preferably at least 50 amino acid residues, and most preferably at least 100 amino acid residues in length. Fragments include chimeric constructs composed of at least a portion of the relevant gene and another molecule. The ability of a candidate fragment to exhibit a biological activity of the anti-angiogenic polypeptide can be assessed by methods known to those skilled in the art, e.g., by its ability to inhibit proliferation of bovine capillary

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cells, or by its ability to inhibit growth of primary tumor cells, e.g., as described herein. See, e.g., Example 9. Also included are fragments containing residues that are not required for biological activity of the fragment or that result from alternative mRNA splicing or alternative protein processing events.

Internal or terminal fragments of a polypeptide can be generated by removing one or more nucleotides from one end (for a terminal fragment) or both ends (for an internal fragment) of a nucleic acid which encodes the polypeptide.

In preferred embodiments, the gene therapy vector of this invention is capable of hybridizing to the native anti-angiogenesis polypeptide-encoding regions and has at least about 80%, preferably at least about 90%, and more preferably at least about 95%, sequence identity to the native nucleotide sequences, and encodes a polypeptide which has anti-angiogenic activity; or a biologically active fragment of any of the above nucleotide sequences wherein the encoded polypeptide has anti-angiogenic activity.

The nucleotide sequences of the present invention can be in the form of RNA or DNA, and the nucleotide sequence can be double-stranded or single stranded and, if single stranded, can be the coding strand or non-coding (antisense) strand.

The coding sequence which encodes the anti-angiogenic polypeptide can be identical to the native coding sequences, or can be a different coding sequence which, as a result of the degeneracy of the genetic code, encodes the same anti-angiogenic polypeptide.

In certain embodiments, the gene therapy vector also has a nucleotide sequence encoding a signal polypeptide and/or export signal (SP) for effecting secretion of the anti-angiogenic polypeptide. Examples of signal polypeptide and/or export signal include plasminogen signal polypeptide and/or export

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signal. Preferably, the signal polypeptide and/or export signal is 5' (i.e., upstream) of the nucleotide sequence encoding the anti-angiogenic polypeptide.

Preferably, the gene therapy vector has a nucleotide sequence encoding a preactivation polypeptide and/or export signal (PAP), which is a small polypeptide and/or export signal which effects folding and secretion of the anti-angiogenic polypeptide *in vivo*. Examples of preactivation polypeptide and/or export signal include plasminogen preactivation polypeptide and/or export signal, described herein, and PAP's of other proteins in the blood clotting cascade.

Preferably, the preactivation polypeptide and/or export signal is positioned 5' of the nucleotide sequence encoding the anti-angiogenic polypeptide. In embodiments which have a signal sequence and an anti-angiogenic polypeptide, preferably the preactivation polypeptide and/or export signal is 5' of the nucleotide sequence encoding the anti-angiogenic polypeptide, and 3' of the nucleotide sequence encoding the signal polypeptide and/or export signal.

We have discovered that results obtained using constructs containing a PAP- encoding nucleic acid sequence are far superior to results using constructs lacking a PAP-encoding sequence. Our hypothesis to explain these unexpectedly superior results with PAP is that, during the complex process by which the anti-angiogenic polypeptide is expressed and processed in living cells, the PAP polypeptide and/or export signal facilitates the export of the polypeptide from the cellular Golgi apparatus and/or the endoplasmic reticulum (ER). The corollary is that, absent PAP, a significant portion of the expressed polypeptide remains trapped in the Golgi and/or ER.

The PAP exemplified herein is derived from human plasminogen; this

-10-

PAP is currently preferred. Our discovery that the use of a PAP dramatically improves results leads us to believe that other PAP's would be useful as well, and such others are therefore contemplated for use in the invention. Thus, as used herein, "PAP" refers to a polypeptide and/or export signal which is naturally associated with a eukaryotic (preferably human) protein, the exportation of which is facilitated by its associated PAP. Examples of other human proteins whose Golgi/ER export is PAP-facilitated include other secreted proteins of the blood coagulation cascade, e.g., fibrinogen, prothrombin, Factor VIII, and Factor IX. Other secreted human proteins also are associated with potentially useful PAPs.

It is not essential that the PAP used in the invention be identical in amino acid sequences to a native PAP; it is well-known that polypeptide and/or export signal that facilitate protein secretion or export, e.g., signal polypeptide and/or export signal and PAPs, can vary from the native forms to a certain extent and still retain their function. Therefore, PAPs useful according to the invention preferably have 75% or greater amino acid sequence identity with a native PAP.

In certain embodiments, the gene therapy vector has a nucleotide sequence encoding a tag for identification of the anti-angiogenic polypeptide and/or export signal. In certain embodiments, the tag is 5' of the nucleotide sequence encoding the anti-angiogenic polypeptide; in other embodiments, the tag is 3' of the nucleotide sequence encoding the anti-angiogenic polypeptide. In embodiments in which the anti-angiogenic polypeptide is endostatin or an angiostatin-endostatin fusion, it is preferred that the tag be 5' of the nucleotide sequence encoding endostatin.

In certain embodiments the gene therapy vector includes a selectable

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marker, e.g., a Neomycin phosphotransferase gene, or a humanized red-shifted green fluorescent protein.

The invention also includes a cell infected or transfected with a gene therapy vector described herein. Preferably, the cell is an animal cell, more preferably an autologous or allogeneic human cell. The gene therapy vectors described herein can be introduced into a cell, e.g., by transformation, transfection, transduction, infection, or ex vivo injection. They can be targeted to a particular cell type.

Administration of nucleic acid, e.g., a gene therapy vector, can be accomplished by any method which allows the nucleic acid to reach the target cells. These methods include, e.g., injection, deposition, implantation, suppositories, oral ingestion, inhalation, topical administration, or any other method of administration where access to the target cells by the nucleic acid is achieved. Injections can be, e.g., intravenous, intradermal, subcutaneous, intramuscular or intraperitoneal. Implantation includes inserting implantable drug delivery systems, e.g., microspheres, hydrogels, polymeric reservoirs, cholesterol matrices, polymeric systems, e.g., matrix erosion and/or diffusion systems and non-polymeric systems, e.g., compressed, fused or partially fused pellets. Suppositories include glycerin suppositories. Oral ingestion doses can be enterically coated. Inhalation includes administering the nucleic acid with an aerosol in an inhalator, either alone or attached to a carrier that can be absorbed.

In certain embodiments of the invention, administration can be designed so as to result in sequential exposures to the nucleic acid over some time period, e.g., hours, days, weeks, months or years. This can be accomplished by repeated administrations of the nucleic acid, e.g., by one of the methods described above, or alternatively, by a controlled release delivery system in

-12-

which the nucleic acid is delivered to the animal over a prolonged period without repeated administrations. By a controlled release delivery system is meant that total release of the nucleic acid does not occur immediately upon administration, but rather is delayed for some time. Release can occur in bursts or it can occur gradually and continuously. Administration of such a system can be, e.g., by long acting oral dosage forms, bolus injections, transdermal patches or subcutaneous implants. Examples of systems in which release occurs in bursts include, e.g., systems in which the nucleic acid is entrapped in liposomes which are encapsulated in a polymer matrix, the liposomes being sensitive to a specific stimulus, e.g., temperature, pH, light, magnetic field, or a degrading enzyme, and systems in which the nucleic acid agent is encapsulated by an ionically-coated microcapsule with a microcapsule core-degrading enzyme. Examples of systems in which release of the nucleic acid is gradual and continuous include, e.g., erosional systems in which the nucleic acid is contained in a form within a matrix, and diffusional systems in which the nucleic acid permeates at a controlled rate, e.g., through a polymer. Such sustained release systems can be, e.g., in the form of pellets or capsules.

The nucleic acid is administered to the patient in a therapeutically effective amount. By therapeutically effective amount is meant that amount which is capable of at least partially preventing or reversing the disease. A therapeutically effective amount can be determined on an individual basis and will be based, at least in part, on consideration of the patient's size, age, the efficacy of the particular nucleic acid used, the type of delivery system used, the time of administration relative to the onset of disease symptoms, and whether a single, multiple, or controlled release dose regimen is employed. A therapeutically effective amount can be determined by one of ordinary skill in the art employing such factors and using no more than routine experimentation.

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In certain embodiments, a therapeutically effective amount of an antiangiogenic polypeptide is administered by providing to the animal a nucleic acid encoding the polypeptide and expressing the polypeptide in vivo. Nucleic acids encoding the polypeptide, or mutants thereof, can be administered in any biologically effective carrier, e.g. any formulation or composition capable of effectively delivering the nucleotide sequence for the anti-angiogenic polypeptide to cells in vivo. Approaches include, e.g., insertion of the nucleic acid into viral vectors. Viral vectors can be delivered to the cells, e.g., by infection or transduction using the virus. Viral vectors can also be delivered to the cells, e.g., by physical means, e.g., by electroporation, lipids, cationic lipids, liposomes, DNA gun, Ca₃(PO₄)₂ precipitation, or delivery of naked DNA. In certain preferred embodiments, the virus is administered by injection, e.g., intramuscular injection, in a dose range of about 10³ to about 10¹⁰ infectious particles per injection, more preferably in a dose range of about 105 to about 108 infectious particles per injection. Single or multiple doses can be administered over a given period of time, depending, e.g., upon the disease.

An alternative is insertion of the nucleic acid encoding the antiangiogenic polypeptide into a bacterial or eukaryotic plasmid. Plasmid DNA can be delivered to cells with the help of, e.g., cationic liposomes (lipofectinTM; Life Technologies, Inc., Gaithersburg, MD) or derivatized (e.g., antibody conjugated) polylysine conjugates, gramacidin S, artificial viral envelopes or other such intracellular carriers, as well as direct injection of the gene construct or $Ca_3(PO_4)_2$ precipitation carried out in vivo, or by use of a gene gun. The above-described methods are known to those skilled in the art and can be performed without undue experimentation.

Since transfer of the nucleic acid to appropriate target cells represents the critical first step in gene therapy, choice of the particular gene delivery

-14-

system will depend on such factors as the intended target and the route of administration, e.g., locally or systemically. Targets for delivery of the nucleic acid can be, e.g., specific target cells which are diseased. For example, the target can be, e.g., the peritoneal cavity, gastro-intestinal tract, bone marrow cavity, liver, lungs, muscles, vasculature, pericardial cavity, pleural cavity, skin, sub-cutaneous or deep connective tissues, central nervous system, spinal fluid, eye, or specific sites of tumor growth. Administration can be directed to one or more cell types, and to one or more cells within a cell type, so as to be therapeutically effective, by methods known to those skilled in the art. For example, the nucleic acid can be, e.g., coupled to an antibody, to a ligand to a cell surface receptor, or to a toxin component, or can be contained in a particle which is selectively internalized into cells, e.g., liposomes, or a virus where the viral receptor binds specifically to a certain cell type, or a viral particle lacking the viral nucleic acid, or can be administered by local injection.

In certain embodiments, the nucleic acid is administered to the patient by introducing *ex vivo* the nucleic acid into cells of the patient, or into syngeneic or allogeneic or xenogeneic cells, and then administering the cells having the nucleic acid to the animal. Any cell type can be used. In certain embodiments, the cells having the introduced nucleic acid are expanded and/or selected after the nucleic acid transfer. The cells having the transferred nucleic acid are subsequently administered to the patient. Preferably, the cells are administered in a dose range of about 1 x 10⁶ to about 1 x 10⁹ cells/dosage/day, and most preferably at about 1 x 10⁷ to about 1 x 10⁸ cells/dosage/day. The cells can be administered by any method which results in delivering the transferred nucleic acid in the cells to the desired target. For example, the cells can be implanted directly into a specific tissue of the patient, or implanted after encapsulation within an artificial polymer matrix. Examples of sites of

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implantation include, e.g., the peritoneal cavity, gastro-intestinal tract, bone marrow cavity, liver, lungs, muscles, vasculature, pericardial cavity, pleural cavity, skin, sub-cutaneous or deep connective tissues, central nervous system, spinal fluid, eye, or specific sites of tumor growth.

Systemic delivery can be achieved, e.g., by introducing the nucleic acid into cells which circulate in the peripheral blood of the patient, or which give rise to cells which circulate in the peripheral blood. In certain embodiments, the nucleic acid is introduced into such cells *ex vivo*, and these cells are then administered to the patient, resulting in systemic delivery within the peripheral blood. These cells can be the cells of the patient or allogeneic cells. Preferred cells in which the nucleic acid can be introduced are hematopoietic cells.

In certain embodiments, other therapy is additionally administered. For example, if the animal is being treated for a tumor, other tumor therapy, e.g., another therapeutic agent, chemotherapy, radiation or surgery, is additionally administered to the patient, either simultaneously or at different times.

Treating is meant to include, e.g., preventing, treating, reducing the symptoms of, or curing the disease. I.e. treating a tumor includes preventing growth of the tumor, causing shrinkage of the tumor, or preventing development of micro-metasteses.

Preferably, the recombinant nucleic acid is a gene therapy vector, e.g., as described herein. Preferably, the anti-angiogenic polypeptide is angiostatin, endostatin, an angiostatin-endostatin fusion protein, or biologically active analogs or fragments thereof. In certain embodiments, the angiostatin has kringles 1, 2 and 3; in other embodiments, the angiostatin has kringles 1, 2, 3 and 4, and, in some embodiments, kringle 5 of human or murine plasminogen. Angiostatin is described in O'Reilly and Folkman U.S. Patent No. 5,639,725, hereby incorporated by reference. Endostatin is described in O'Reilly and

Folkman PCT Appln. No. WO 97/15666, published May 1, 1997, hereby incorporated by reference.

In certain embodiments, the recombinant nucleic acid has been introduced ex vivo into cells so as to express the anti-angiogenic polypeptide in the cells, and the recombinant nucleic acid is administered to the patient by administering to the patient the cells containing the recombinant nucleic acid. In certain embodiments, the cells are derived from the patient; in other embodiments the cells are allogeneic cells relative to the cells of the patient.

Where cells are infected or transfected *ex vivo* for later infusion into the patient, the cells are preferably hematopoietic cells, but can also be mesenchymal cells, stem cells, epithelial cells (e.g., from the gut), or dendritic cells.

The gene therapy vectors of the invention can be provided in a pharmaceutical composition comprising a therapeutically effective amount of the recombinant nucleic acid together with a pharmaceutically acceptable carrier. Pharmaceutically acceptable carriers include, e.g., water, saline, dextrose, glycerol, ethanol, liposomes and lipid emulsions.

The following non-limiting examples further illustrate the present invention.

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EXAMPLES

Example 1: Construction of Inserts for Gene Therapy Vectors Containing cDNA for Angiostatin, Endostatin or Angiostatin-Endostatin Fusion Proteins

The following genetic constructs are inserted into retroviral gene therapy vectors; the genetic constructs contain human or murine cDNA for angiostatin, endostatin or an angiostatin-endostatin fusion, and DNA encoding a signal

polypeptide and/or export signal (SP), a tag (FLAG), and, preferably, a preactivation polypeptide and/or export signal (PAP). The constructs are all made using standard genetic engineering techniques, and their insertion into retroviral gene therapy vectors is carried out using known methods. The constructs have the following components:

	Murine Constructs	
	SP-K1-K2-K3-Flag	
	SP-K1-K2-K3-K4-Flag	
	SP-K1-K2-K3-K4-K5-Flag	
10	SP-PAP-K1-K2-K3-Flag	
	SP-PAP-K1-K2-K3-K4-Flag	(SEQ ID NO: 11 and 12)
	SP-Flag-Endo	(SEQ ID NO: 13 and 14)
	SP-K1-K2-K3- Flag-Endo	
	SP-K1-K2-K3-K4- Flag-Endo	(SEQ ID NO: 15 and 16)
15	SP-PAP-K1-K2-K3- Flag-Endo	
	Human Constructs	
	SP-K1-K2-K3	
	SP-K1-K2-K3-K4	
	~~ *** *** ***	
	SP-K1-K2-K3-K4-K5	
20	SP-R1-K2-K3-K4-K5 SP-PAP-K1-K2-K3	
20		
20	SP-PAP-K1-K2-K3	
20	SP-PAP-K1-K2-K3 SP-PAP-K1-K2-K3-K4	
20	SP-PAP-K1-K2-K3 SP-PAP-K1-K2-K3-K4 SP-PAP-K1-K2-K3-K4-K5	

-18-

Nucleic acid and amino acid sequences for mouse and human angiostatin and endostatin used in these constructs are shown in Figs. 5-7.

Nucleic acid and amino acid sequence of the FLAG peptide:

amino terminus-5'- ASP TYR LYS ASP ASP ASP LYS GAC TAC AAG GAC GAC GAT GAC AAG

Human plasminogen derivative constructs

The entire coding region of the human plasminogen cDNA from the start (ATG) to the stop (TAA) codon is 2433bp in size.

This sequence encodes a signal peptide (bp 1-57), a preactivation peptide (bp 58-288), and 5 distinct structural regions known as kringles (K1-K3 from bp 289-1092; K4 from bp1093-1380; K5 from bp 1381-1740). Please note that although I have given precise bp measurements for kringles K4 and K5, it can be argued that the sequence encoding K4 is between bp1056-1440 and the sequence encoding K5 is between bp1362-1680.

A DNA fragment encoding a portion of the human plasminogen protein from bp 1 to 1377 was obtained by PCR of a widely available human liver cDNA library using synthetic DNA oligonucleotides complementary to sequences immediately preceding the signal peptide and immediatly following kringle 4. This fragment contains the sigal peptide (bp1-57), the preactivation peptide (bp 58-288), kringles 1 (bp289-549), 2 (bp 550-804), 3 (bp 805-1092) and 4 (bp 1093-1380). The synthetic oligonucleotides used for this reaction contained engineered recognition sites for the restriction enzymes EcoRI and XhoI. Following the PCR reaction the amplified fragment was closed into the EcoRI/XhoI sites of BluescriptSK(-) (Stratgene) using standard techniques (Maniatis). Following cloning the integrity of the amplified sequence was verified by sequencing both strands using the Sanger method (Sanger). Various derivatives of the cloned fragment were subsequently constructed using BluescriptSK(-) (Stratagene) as a backbone. A full list of the derivatives are described in Table 1. Briefly, the variations are composed of constructs containing various combinations of kringles with or without the signal and/or preactivation peptide sequences. These derivatives were constructed using both standard techniques as well as PCR and the use of double stranded synthetic oligonucleotides. In all cases the integrity of the start codon, coding sequence and termination codon was verified by double stranded sequencing using the Sanger method.

Murine plasminogen derivative constructs

The coding sequence for murine plasminogen is 2439bp in size and, similar to the human plasminogen cDNA encompasses a sequence encoding signal and preactivation peptides (bp 1-57 and 58-288 respectively) in addition to 5 kringle regions; kringle 1-3 (bp 289-1092), kringle 4 (bp 1093-1380) and kringle 5 (bp 1381-1743). Again, although I have given precise bp measurements for kringles K4 and K5, it can be argued that the sequence encoding K4 is between bp1056-1440 and the sequence encoding K5 is between bp1362-1680.

The murine plasminogen cDNA has previously been cloned and was made available to us. Derivatives of murine plasminogen were constructed using sequences derived from bp 1-1743 of the coding sequence. Various combinations of kringle regions with or without signal and preactivation peptide regions were made using BluescriptSK(-) (Stratagene, La Jolla, CA) as the vector backbone. These derivatives were constructed using standard cloning techniques (Maniatis, Molecular cloning; a laboratory manual, second edition, 1989) in combination with PCR utilizing synthetic oligonuleotides using

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Angiostatin function was not altered by adding the FLAG polypeptide and/or export signal to either the N- or C-terminal ends, whereas endostatin was functional only if FLAG was added to its N-terminal end.

Example 2: Construction of Retroviral Gene Therapy Vectors

This example illustrates the construction of retroviral gene therapy vectors comprising cDNA for angiostatin, endostatin or angiostatin-endostatin fusion proteins.

The DNA inserts from Example 1 were inserted into two retroviral vectors. Both vectors were derived from the Murine Stem Cell Virus (MSCV), which is a variant of Moloney Murine Leukemia Virus (MoMLV) having several mutations allowing high, sustained expression in hematopoietic stem cells and their progeny. In both cases, the angiostatin, endostatin, or angiostatin-endostatin fusion DNA inserts were under the transcriptional control of the retroviral left Long Terminal Repeat (LTR). In the first vector, the dominant selectable marker was the Neomycin phosphotransferase gene (NeoR), which confers resistance to G418, and is driven by an internal phosphoglycerate kinase (PGK) promoter. In the second vector, the dominant selectable marker was the humanized, red-shifted green fluorescent protein (EGFP), which is co-translationally expressed by means of an Internal Ribosome Entry Site (IRES) from the Encephalomyocarditis virus (EMCV).

The retroviral gene therapy vectors were transfected by CaPO₄ precipitation in the transient ecotropic packaging cell-line BOSC 23, Pear et al., *PNAS* <u>90</u>:8392 (1993). Viral supernatants were collected two days thereafter and filtered through 0.45 mm filters. Filtered viral supernatants were subsequently used to infect GENETIX's stable amphotropic retroviral packaging cell-line AM12 (Genetix Pharmaceuticals, Inc., Cambridge, MA). After another two days, viral supernatants from transduced AM12 were filtered

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and used to infect GENETIX's stable ecotropic retroviral packaging cell-line GP+E86 (Genetix Pharmaceuticals, Inc.). Both transduced AM12 and GP+E86 were then selected in the presence of G418 (in the case of constructs bearing NeoR) or sorted by Fluorescent Activated Cell Sorter (FACS) for EGFP expression. Viral titers were estimated according to standard practice by counting G418 resistant colonies among NIH3T3 cells exposed to diluted virus preparation. Ecotropic viral titers were above 5 x 10⁵ /ml of viral supernatants, only 3-fold lower than "empty" control vectors. No Replication Competent Retrovirus (RCR) was detected in standard assays.

10 Example 3: Transduction of Target Cells Using Retroviral Gene Therapy Vectors

This example illustrates the stability of retroviral gene therapy vector transmission and the lack of toxicity in non-endothelial target cells.

Following 24-hour incubation of confluent viral producer cells in 100 mm plates, viral supernatant was removed and filtered (0.45 µm filter, Gelman Sciences, Ann Arbor, MI). Viral supernatant, containing 7 µg/ml polybrene (Sigma, St. Louis, MO), was added to target cells 24 hours after plating the target cells. Fresh medium was added after 4-12 hours, and, after an additional 48 hours, cells were selected for retroviral infection by exposure to medium containing 1 mg/ml G418 (Gibco BRL, Grand Island, NY) or by FACS sorting (FACStar cell sorter, Becton Dickinson, San Jose, CA). The stability of transmission of the retroviral gene therapy vectors described in Example 2 was examined by Southern blot analysis of transduced NIH3T3 cells, using specific probes (EGFP) and restriction enzyme digestion of genomic DNA with Sac1, which cuts only once in each LTR. Stable chromosomal integration of intact proviruses of appropriate length was observed with all constructs.

The lack of non-specific toxicity on non-endothelial cells was

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established by using filtered viral supernatants to transduce various tumor cell-types and cell-lines (NIH3T3 cells, K562 cells (ATCC), and human SK-N-AS neuroblastoma cells; Cohen, P.S., *Cancer Research*, <u>55</u>:2380 (1995).

Transduced cell populations were subsequently selected with G418 or sorted for EGFP expression by FACS. No obvious effects on cell viability, growth or other phenotypical characteristics were detected.

Example 4: Protein Expression of Angiostatin, Endostatin and Angiostatin-Endostatin Fusion Proteins in Cells Transduced with Retroviral Gene Therapy Vectors

This example illustrates that recombinant angiostatin, endostatin, and angiostatin-endostatin fusion proteins were readily detected in retrovirally transduced cells and their supernatant, indicating efficient expression and secretion.

MSCV virus based vectors containing sequences encoding murine Kringle 1 (K1), K1K5, K1K2K3, K1K2K3K4, and K1K2K3K4K5 were used to transduce NIH3T3 cells. With regard to the murine recombinant proteins, Western blot analysis of transduced cells and their supernatant was performed by means of a monoclonal antibody that recognizes the FLAG polypeptide and/or export signal. Because this antibody is not mono-specific, significant cross-reactivity with murine proteins was apparent. However, by comparing the pattern obtained with mock cells, it was clear that the antibody revealed an additional band of appropriate size in all transduced cells. Moreover, the recombinant proteins were detected in cell supernatants at levels above 50 ng/ml, using a protein concentration/semi-purification procedure (Centricon columns, Amicon, Beverly, MA). With regard to the human recombinant proteins, no FLAG tag was added, so a monoclonal antibody that recognizes specifically the first three kringles of human plasminogen in its native, non-

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denatured form was used; O'Reilly et al., *Cell* 79:315 (1994). Because of this constraint, Western blot analysis using denaturing gels could not be performed. An ELISA assay was performed which indicated that human recombinant angiostatin was detected at levels likely to be therapeutic according to previous findings in the model of Lewis Lung Carcinoma *Id*.

These results indicate that high levels of recombinant proteins of expected length were expressed in retrovirally transduced cells and were efficiently secreted.

Example 5: In Vivo Anti-Tumor Activity of Cells Transduced with Gene
Therapy Vectors Encoding the Angiostatin-Endostatin Fusion
Protein

Human SK-N-AS neuroblastoma cells (Cohen, 1995) were transduced with the retroviral gene therapy vector containing the angiostatin-endostatin fusion protein, described in Example 2. These cells (1,000,000) were suspended in 1 mL Dulbecco's phosphate buffered saline and injected into the right mid-quadrant of nude immuno-compromised mice. While no impairment of the in vitro growth of transduced cells was observed, a dramatic decrease in tumor growth in nude mice cells following subcutaneous implantation of the transduced cells was evident as compared to "mock virus"-transduced control cells.

Ex Vivo Transfer of Retroviral Gene Therapy Vectors Encoding Anti-Angiogenic Polypeptides to Primary Hematopoietic Cells, and Subsequent Transplantation to Recipient Mice

This example illustrates infection of primary hematopoietic cells from

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donor mice with retroviral gene therapy vectors encoding angiostatin, endostatin, or an angiostatin-endostatin fusion protein, and a selectable GFP marker, and subsequent transplantation of the transduced hematopoietic cells into recipient mice.

Femoral bone marrow cells are harvested from male donor C57BL6/J-Ly5.1 mice (Jackson Labs, Bar Harbor, ME), intravenously injected four days previously with 150 mg/kg of 5-fluorouracil (5-FU). Bone marrow cells are cultured for two days in medium composed of DMEM, 15% fetal calf serum, 10 ng/ml human IL-6, 6 ng/ml murine IL-3 and 100 ng/ml murine Steel factor prior to two days of culture atop a confluent monolayer of irradiated (1,500 cGy, ¹³⁷Cs γ-irradiation) viral producer cells in the above medium including 6 ug/ml of prolamine sulfate. The viral producer cells are transfected with a retroviral gene therapy vector, as described above. Upon completion of the coculture infection protocol, recovered non-adherent cells are cultured for an additional 48 hours to allow for expression of the transferred GFP gene. Retrovirally transduced cells expressing the transferred GFP gene are subsequently identified and selected for, using a FACStar+ cell sorter (Becton Dickinson, San Jose, CA). The GFP+ cells are intravenously injected into congenic female C57BL6/J-Ly5.2 recipient mice (National Cancer Institute, Washington, DC) previously given 950 cGy (83cGy/min, ¹³⁷Cs γ-rays) of whole body irradiation. In each case, a small fraction of GFP+ sorted cells is used for day 12 CFU-S and in vitro clonogenic progenitor assays to assess the efficiency of the infection and selection procedures on these more mature cell types.

25 Example 7: Engraftment of Recipient Mice with Donor-Derived Hematopoietic Cells

This example illustrates engraftment of the recipient mice with the

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donor-derived transfected hematopoietic cells from Example 6.

The donor and recipient mice are phenotypically distinguishable on the basis of Y chromosome specific sequences, as well as on the basis of allelic differences at the murine CD45 cell surface antigen locus. Male donor mice are homozygous for the CD45.2 allele, while female recipient mice are homozygous for CD45.1. The engraftment of recipient mice with donor-derived (CD45.2+) cells is assessed at both short (5 weeks) and long (34 months) time points post-transplant by flow cytometric analysis of peripheral blood samples stained with a phycoerythrin labeled antibody specific for the CD45.2 antigen (Pharmingen, San Diego, CA). The results indicate that engraftment occurs.

Example 8: Proviral Marking and GFP Expression in Recipient Mice

This example illustrates the presence of recombinant provirus and expression of the transferred GFP gene in the recipient mice from Example 6.

The level of proviral marking in reconstituted animals is initially determined by Southern blot and semi-quantitative PCR analysis of DNA obtained from peripheral blood leukocytes. The large majority of donorderived (CD45.2+) cells in recipient mice contain a minimum of one copy of recombinant provirus. In addition, flow cytometric analysis of peripheral blood leukocytes is performed to ascertain the proportion of cells expressing the transferred GFP cDNA. Because the GFP and angiogenic inhibitor protein cDNAs are both driven from the same regulatory sequences, due to the inclusion of an internal ribosomal entry site (IRES) element, the analysis of GFP expression in the peripheral blood provides an indirect measurement of the levels of anti-angiogenic protein being expressed. The results indicate expression of the transferred genes.

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Example 9: Anti-Angiogenic Polypeptide Expression in Recipient Mice

This example illustrates the presence of anti-angiogenic polypeptide in the sera of the recipient mice from Example 6, using both physical and functional assays.

Serum obtained from the transplanted animals described in Example 6 is used for ELISA using an antibody specific for the synthetic FLAG epitope (IBI, Eastman Kodak, New Haven, CT) and compared against known standards of purified protein. Results indicate the presence of the anti-angiogenic polypeptide in the serum.

To determine whether a functional anti-angiogenesis polypeptide is present in the circulation, sera from transplanted animals is tested for its ability to inhibit the proliferation of bovine capillary cells *in vitro*; O'Reilly (1994). Briefly, cells are plated in 24 well dishes at 25,000 cells/ml and maintained in DMEM with 5% bovine calf serum for 24 hours. The medium is then replaced with fresh medium containing various dilutions of the test serum. After 20 minutes of incubation, fresh medium including b-FGF (final concentration 1 ng/ml) is added and the cells are cultured for 72 hours. Cells are then dispersed using trypsin and the cell number determined by Coulter counter. Results indicate that functional anti-angiogenic polypeptide is present in the sera of the recipient mice.

In addition, the ability of circulating anti-angiogenic polypeptide to inhibit the growth of primary tumor cells is assessed. Transplanted mice are subcutaneously injected with one million Lewis lung carcinoma (LLC) cells (O'Reilly, (1994)) at the proximal midline of their dorsal skin. The mice are closely monitored for survival, tumor size and growth, and overall health. Results indicate that the anti-angiogenic polypeptides from the sera of the

-26-

recipient mice inhibit growth of the LLC tumor cells.

Finally, upon sacrifice of the transplanted recipient mice, blood, spleen, thymus and bone marrow are harvested and analyzed for the presence of proviral DNA by Southern analysis as well as expression of the transferred GFP and anti-angiogenic polypeptide cDNAs by flow cytometry and ELISA. Moreover, a portion of bone marrow cells is re-transplanted into secondary recipients to generate individual day 12 spleen colonies, as well as plated in methylcellulose to assess in vitro clonogenic progenitors. Individual clones are analyzed for proviral DNA by PCR or Southern blot, and for gene expression by flow cytometry and ELISA. Results of these tests also indicate the presence of proviral DNA and expression of the anti-angiogenic polypeptides and marker proteins.

Example 10: Evaluating the Efficacy of Retroviral Gene Therapy Vectors
Encoding Anti-Angiogenic Polypetides on Various Human
Cancers Implanted in SCID Mice Using Ex Vivo Gene Therapy

This example illustrates a method for rapidly screening various forms of human cancer to determine susceptibility to treatment by the systemic delivery of anti-angiogenic polypeptides.

The methods for gene transfer, assessment of proviral marking and assessment of transferred gene expression as described in Examples 3 through 9 are repeated using immuno-deficient SCID mice, with the following exceptions. Since SCID mice are more sensitive to γ-irradiation than C57BL6/J mice, the female SCID recipients receive a lower dose of 400cGy of whole body irradiation in contrast to the 950cGy required for C57BL6/J. In addition, since the SCID mice do not possess allelic differences at the CD45 cell surface antigen locus, donor and recipient cells are phenotypically distinguished on the basis of Y chromosome specific sequences using Southern

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blot analysis.

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Bone marrow from male donor SCID mice is infected, selected for on the basis of expression of the transferred GFP marker cDNA, and transplanted into irradiated female SCID recipients. Engraftment with provirally marked cells and expression of the transferred genes is demonstrated. The mice are then separately implanted with a variety of human tumor cell types, e.g., breast adenocarcinoma, lung squamous cell carcinoma, and brain glioblastoma. In each case, the ability of the anti-angiogenic polypeptides to inhibit the growth of the various human tumor cell types is monitored and quantified.

10 Example 11: Evaluating the Efficacy of Retroviral Gene Therapy Vectors

Encoding Anti-Angiogenic Polypeptides for Treatment of

Ovarian Cancer Using In Vivo Gene Therapy

This example illustrates the feasibility of using retroviral gene therapy vectors encoding anti-angiogenic polypeptides to achieve efficient gene transfer to established tumors <u>in vivo</u> using a well-established murine model of human ovarian cancer. Following injections, mice are closely monitored for tumor growth and survival.

Eight to ten week old nude mice (Jackson Labs, Bar Harbor are injected intra-peritoneally with 1 x 10⁷ PA-1 cells, an ovarian cancer cell-line (ATCC), and followed until palpable tumors are identified. Viral supernatant for in vivo injection is prepared as follows: Viral producer cells are grown to confluence in DMEM with 10% bovine calf serum, and the medium is then changed. After 24 hours of incubation, the viral conditioned supernatant is filtered though a 0.45 um low protein binding filter, protamine sulfate is added to a final concentration of 6ug/ml, the solution is aliquoted into 2 ml volumes, and frozen at -80°C. Recipient mice receive three intraperitoneal injections of viral supernatant (2 mls per injection) in addition to the polycation, over a period of

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36 hours. Control mice are injected with medium collected from confluent dishes of NIH3T3 cells. Following injection of the viral conditioned supernatant, the mice are analyzed for survival as well as tumor growth over time as compared to mock injected controls. Results indicate that treatment of the ovarian cancer occurs. At death, the tumors are removed, weighed, and the cells dissociated for DNA extraction for Southern blot analysis to detect recombinant provirus.

Those skilled in the art will be able to ascertain using no more than routine experimentation, many equivalents of the specific embodiments of the invention described herein. These and all other equivalents are intended to be encompassed by the following claims.

In other embodiments, the invention provides methods and compositions for treating diseases and processes that are mediated by angiogenesis including, but not limited to, hemangioma, solid tumors, leukemia, metastasis, telangiectasia, psoriasis, scleroderma, pyogenic granuloma, myocardial angiogenesis, plaque neovascularization, coronary collaterals, cerebral collaterals, arteriovenous malformations, ischemic limb angiogenesis, corneal diseases, rubeosis, neovascular glaucoma, diabetic retinopathy, retrolental fibroplasia, arthritis, diabetic neovascularization, macular degeneration, wound healing, peptic ulcer, *Helicobacter* related diseases, fractures, keloids, vasculogenesis, hematopoiesis, ovulation, menstruation, placentation, and cat scratch fever.

What is claimed is:

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CLAIMS

- 1. Use of a nucleic acid molecule which expresses an anti-angiogenic polypeptide selected from the group consisting of human angiostatin, murine angiostatin, human endostatin, murine endostatin, and angiogenesis-inhibiting fragments thereof in the preparation of a medicament for inhibiting tumor growth in a human patient harboring a solid tumor, wherein expression of the anti-angiogenic polypeptide in the patient inhibits angiogenesis in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells, thereby inhibiting its growth.
- 2. Use of a nucleic acid molecule which expresses an anti-angiogenic polypeptide of the formula A-B, wherein

A and B are polypeptide and/or export signal joined by a polypeptide and/or export signal bond;

A contains at least amino acids and comprises kringles 1, 2, and 3 of human or murine angiostatin; and

B contains at least amino acids and includes at least 75% of the amino acid sequence of human or murine endostatin in the preparation of a medicament for inhibiting tumor growth in a human patient harboring a solid tumor, wherein expression of the anti-angiogenic polypeptide in the patient inhibits angiogenesis in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells, thereby inhibiting its growth.

3. The use of claim 2, wherein A further comprises kringle region 4 of

-30-

human or murine angiostatin.

- 4. The use of claim 2 or claim 3, wherein A further comprises kringle 5 of human or murine plasminogen.
- 5. The use of claim 1 or claim 2, wherein said nucleic acid moleculeconstitutes a portion of a viral vector.
 - 6. The use of claim 1 or claim 2, wherein said nucleic acid molecule constitutes a portion of a plasmid.
 - 7. The use of claim 6, wherein said plasmid is carried in a cell-free carrier so that the plasmid transfects living cells of the patient following plasmid administration, causing expression of the anti-angiogenesis polypeptide and/or export signal in the patient such that angiogenesis in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells is inhibited, causing inhibition of tumor growth.
- 8. The use of claim 6, wherein said plasmid has been transfected into animal cells ex vivo, wherein said animal cells express the anti-angiogenesis polypeptide to inhibit tumor-associated angiogenesis and tumor growth.
- 9. The use of claim 5, wherein said viral vector is carried in a cell-free carrier, so that the viral vector is incorporated into living cells of the patient
 20 following viral vector administration, causing expression of the anti-

angiogenesis polypeptide in the patient such that angiogenesis in the vicinity of the tumor and/or systemically by diffusion of the recombinant protein to the vascular compartment from secreting transduced cells is inhibited, causing inhibition of tumor growth.

- The use of claim 5, wherein animal cells are infected with said viral vector *ex vivo* and then administered to said patient, wherein said animal cells express the anti-angiogenesis polypeptide to inhibit tumor-associated angiogenesis and tumor growth.
 - 11. The use of claim 8, wherein said animal cells are human cells.
 - 12. The use of claim 11, wherein said human cells are autologous.
 - 13. The use of claim 11, wherein said human cells are allogeneic.
 - 14. The use of claim 10, wherein said animal cells are human cells.
 - 15. The use of claim 14, wherein said human cells are autologous.
 - 16. The use of claim 14, wherein said human cells are allogeneic.
- 15 17. The use of claim 5, wherein said viral vector is a retroviral vector.
 - 18. The use of claim 5, wherein said viral vector is a non-retroviral vector selected from the group consisting of adenoviral, adeno-associated, herpes, Simliki Forest virus, and poxvirus vectors.

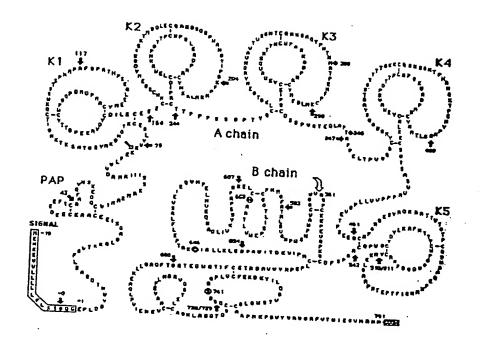
-32-

- 19. The use of claim 17, wherein said retroviral vector is Murine Stem Cell Virus or a lentivirus.
- 20. The use of claim 1, wherein said angiostatin comprises kringles 1, 2 and 3.
- 5 21. The use of claim 20, wherein said angiostatin further comprises kringle 4.
 - 22. The use of claim 1, wherein said anti-angiogenic polypeptide is a fusion of angiostatin or a biologically active fragment thereof and endostatin or a biologically active fragment thereof.
- 23. The use of claim 1, wherein said nucleic acid molecule includes a nucleotide sequence encoding a signal polypeptide and/or export signal for effecting secretion of said anti-angiogenesis polypeptide.
 - 24. The use of claim 23, wherein said signal polypeptide and/or export signal is plasminogen signal polypeptide and/or export signal.
- 15 25. The use of claim 1, wherein said nucleic acid molecule includes a nucleotide sequence encoding a preactivation polypeptide and/or export signal for effecting Golgi and/or ER export of the anti-angiogenic polypeptide..
 - 26. The use of claim 25, wherein said preactivation polypeptide and/or export signal is a preactivation polypeptide and/or export signal of a human protein of the blood coagulation cascade.

- 27. The use of claim 26, wherein said preactivation polypeptide and/or export signal is human plasminogen preactivation polypeptide and/or export signal.
- 28. The method of claim 25, wherein the preactivation encoding sequence is positioned between a signal-encoding sequence and the sequence encoding the anti-angiogenic polypeptide and/or export signal.
 - 29. The use of claim 1, wherein said nucleic acid molecule includes a nucleotide sequence encoding a tag for identification of said anti-angiogenic polypeptide.
 - 30. The method of claim 27, wherein said tag is a Flag tag polypeptide and/or export signal.
 - 31. A viral gene therapy vector comprising a nucleic acid molecule which encodes an anti-angiogenic polypeptide selected from the group consisting of human angiostatin, murine angiostatin, human endostatin, murine endostatin, and angiogenesis-inhibiting fusions and fragments thereof, wherein said viral vector is sufficiently attenuated for use in human gene therapy.
 - 32. A human cell infected with the vector of claim 31.
- 33. Use of a nucleic acid molecule which expresses in said patient an anti-angiogenic polypeptide selected from the group consisting of human
 angiostatin, murine angiostatin, human endostatin, murine endostatin, and angiogenesis-inhibiting fusions and fragments thereof, in the preparation of a

medicament for treating a human patient suffering from diabetic retinopathy, wherein expression of the anti-angiogenic polypeptide in the patient inhibits angiogenesis in the vicinity of the retina.

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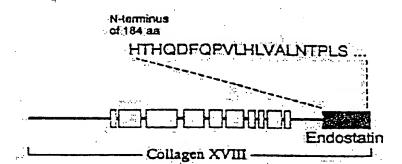
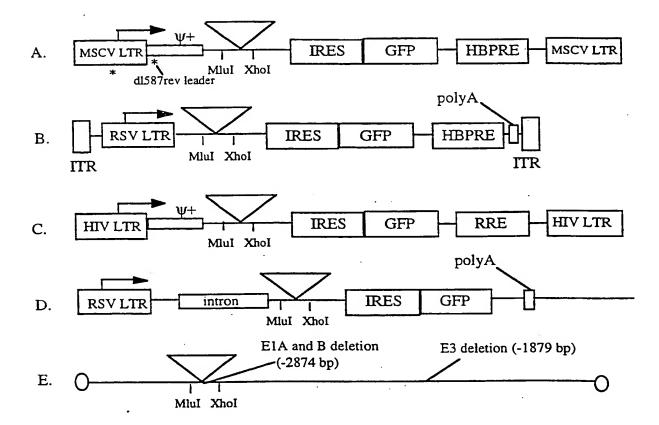
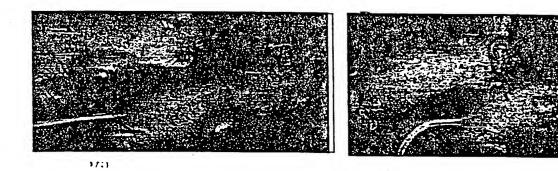


Fig. 2





2410 2420 2430 ACTTGGATTGACCGACTGATGAGAAATAATTAA ThrTTplleGluGlyValMetArgAsnAsn

RNSDOCID: <WO 9926480A1 L >

Pend human plasminogen coding sequence.

Fig 6

Atabal radio Atara sanga Atara

CACCCCCCCC PruProPro

900 CCCCTGGAGT ArgatypSer

EUU CTCCANCACC SerlysThr

SCO GETACACTAC Tyr The The

TIKICCCCACI TIKICCCCACI

National Author

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THEIR USE IN TREATING ANGIOGENISIS-RELATED DISEASES

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ctg ttc agt Leu Phe Ser 35	gtc act aag aag cag ctg gga gca gga agt ata gaa gaa 14 Val Thr Lys Lys Gln Leu Gly Ala Gly Ser Ile Glu Glu 40 45	4
tgt gca gca Cys Ala Ala 50	a aaa tgt gag gag gac gaa gaa ttc acc tgc agg gca ttc 19 a Lys Cys Glu Glu Asp Glu Glu Phe Thr Cys Arg Ala Phe 55 60	2
caa tat cac Gln Tyr His 65	c agt aaa gag caa caa tgt gtg ata atg gct gaa aac agg 24 s Ser Lys Glu Gln Gln Cys Val Ile Met Ala Glu Asn Arg 70 75 80	0
aag too too Lys Ser Ser	c ata atc att agg atg aga gat gta gtt tta ttt gaa aag 28 r Ile Ile Ile Arg Met Arg Asp Val Val Leu Phe Glu Lys 85 90 95	8

DNEDOCID- JWO GOSEAROAT I -

														Tyr	aga Arg	336
															agt Ser	384
tcc Ser	act Thr 130	tct Ser	ccc Pro	cac His	aga Arg	cct Pro 135	aga Arg	ttc Phe	tca Ser	cct Pro	gct Ala 140	aca Thr	cac His	ccc Pro	tca Ser	432
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Asp	Ile	Leu	Glu 180	Cys	gaa Glu	Glu	Glu	Cys 185	Met	His	Cys	Ser	Gly 190	Glu	Asn	576
Tyr	Asp	Gly 195	Lys	Ile	tcc Ser	Lys	Thr 200	Met	Ser	Gly	Leu	Glu 205	Cys	Gln	Ala	624
Trp	Asp 210	Ser	Gln	Ser	cca Pro	His 215	Ala	His	Gly	Tyr	Ile 220	Pro	Ser	Lys	Phe	672
Pro 225	Asn	Lys	Asn	Leu	aag Lys 230	Lys	Asn	Tyr	Cys	Arg 235	Asn	Pro	Asp	Arg	Glu 240	720
Leu	Arg	Pro	Trp	Cys 245	ttc Phe	Thr	Thr	Asp	Pro 250	Asn	Lys	Arg	Trp	Glu 255	Leu	768
Cys	Asp	Ile	Pro 260	Arg	tgc Cys	Thr	Thr	Pro 265	Pro	Pro	Ser	Ser	Gly 270	Pro	Thr	816
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	_										_	aag Lys			1200
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Thr Asp Ser Cys Gln Gly Asp Ser Gly Gly Pro Leu Val Cys Phe Glu
755

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Lys Asp Lys Tyr Ile Leu Gln Gly Val Thr Ser Trp Gly Leu Gly Cys
770

gca cgc ccc aat aag cct ggt gtc tat gtt cgt gtt tca agg ttt gtt
Ala Arg Pro Asn Lys Pro Gly Val Tyr Val Arg Val Ser Arg Phe Val
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<212> PRT

<213> Homo sapiens

<400> 2

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Cys Asp Ile Pro Arg Cys Thr Thr Pro Pro Pro Ser Ser Gly Pro Thr

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			Asn	645					650					655	_
			Glu 660					665					670		
		675	Val				680					685			
FIO	690	TAT	Val	vaı	MIG	695	Arg	IIII	GIU	cys	700	тте	ınr	σтλ	Trp

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Thr	Asp	Ser 755		Gln	Gly	Asp	Ser 760		Gly	Pro	Leu	Val 765		Phe	Glu	
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785	_	•		Lys	790			-		Arg 795	Val	Ser	Arg	Phe	Val 800	
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Gly	Gln	Gly	Asp 20	Ser	Leu	Asp	Gly	Tyr 25	Ile	Ser	Thr	Gln	Gly 30	Ala	Ser	
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Бец	FILE	35	Deu	1111	БуБ	шуБ	40	Dea	ALU	niu	O ₁	45	vuı	JUL	710p	
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				aaa												240
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Lys	Thr	Ser	Ser	Ile 85	Ile	Arg	Met	Arg	Asp 90	Val	Ile	Leu	Phe	95	Lys	
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J			100			•	-	105	•		•		110	-		
gga	acc	atg	tcc	agg	aca	aag	agt	ggt	gtt	gcc	tgt	caa	aag,	tgg	ggt	384
				Arg												

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		115					120					125	i			
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gag Glu 145	GIY	cta Leu	gaa Glu	gag Glu	aac Asn 150	Tyr	tgt Cys	agg Arg	aac Asn	cca Pro 155	Asp	aat Asn	gat Asp	gaa Glu	Caa Gln 160	480
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													gly aaa			1152
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													gag Glu			1440
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													ctg Leu			1584
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		cct Pro															1776
		tgg Trp 595	_	_	_		_		_								1824
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		gga Gly								_		-		_			2160
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		aag Lys								_							2352

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Tyr Gln Cys Leu Lys Gly Arg Gly Glu Asn Tyr Arg Gly Thr Val Ser 275 280 285 Val Thr Val Ser Gly Lys Thr Cys Gln Arg Trp Ser Glu Gln Thr Pro 295 300 His Arg His Asn Arg Thr Pro Glu Asn Phe Pro Cys Lys Asn Leu Glu 310 315 Glu Asn Tyr Cys Arg Asn Pro Asp Gly Glu Thr Ala Pro Trp Cys Tyr 325 330 Thr Thr Asp Ser Gln Leu Arg Trp Glu Tyr Cys Glu Ile Pro Ser Cys 340 345. Glu Ser Ser Ala Ser Pro Asp Gln Ser Asp Ser Ser Val Pro Pro Glu 360 365 Glu Gln Thr Pro Val Val Gln Glu Cys Tyr Gln Ser Asp Gly Gln Ser 375 380 Tyr Arg Gly Thr Ser Ser Thr Thr Ile Thr Gly Lys Lys Cys Gln Ser 390 395 Trp Ala Ala Met Phe Pro His Arg His Ser Lys Thr Pro Glu Asn Phe 405 410 Pro Asp Ala Gly Leu Glu Met Asn Tyr Cys Arg Asn Pro Asp Gly Asp 425 Lys Gly Pro Trp Cys Tyr Thr Thr Asp Pro Ser Val Arg Trp Glu Tyr 440 Cys Asn Leu Lys Arg Cys Ser Glu Thr Gly Gly Ser Val Val Glu Leu 455 Pro Thr Val Ser Gln Glu Pro Ser Gly Pro Ser Asp Ser Glu Thr Asp 470 475 Cys Met Tyr Gly Asn Gly Lys Asp Tyr Arg Gly Lys Thr Ala Val Thr 485 490 Ala Ala Gly Thr Pro Cys Gln Gly Trp Ala Ala Gln Glu Pro His Arg 500 505 His Ser Ile Phe Thr Pro Gln Thr Asn Pro Arg Ala Asp Leu Glu Lys 515 520 525 Asn Tyr Cys Arg Asn Pro Asp Gly Asp Val Asn Gly Pro Trp Cys Tyr 535 540 Thr Thr Asn Pro Arg Lys Leu Tyr Asp Tyr Cys Asp Ile Pro Leu Cys 550 555 Ala Ser Ala Ser Ser Phe Glu Cys Gly Lys Pro Gln Val Glu Pro Lys 565 570 Lys Cys Pro Gly Arg Val Val Gly Cys Val Ala Asn Pro His Ser 585 Trp Pro Trp Gln Ile Ser Leu Arg Thr Arg Phe Thr Gly Gln His Phe 600 Cys Gly Gly Thr Leu Ile Ala Pro Glu Trp Val Leu Thr Ala Ala His 615 Cys Leu Glu Lys Ser Ser Arg Pro Glu Phe Tyr Lys Val Ile Leu Gly 630 635 Ala His Glu Glu Tyr Ile Arg Gly Leu Asp Val Gln Glu Ile Ser Val 645 650 Ala Lys Leu Ile Leu Glu Pro Asn Asn Arg Asp Ile Ala Leu Leu Lys 660 665 Leu Ser Arg Pro Ala Thr Ile Thr Asp Lys Val Ile Pro Ala Cys Leu 675 680 Pro Ser Pro Asn Tyr Met Val Ala Asp Arg Thr Ile Cys Tyr Ile Thr . 695 Gly Trp Gly Glu Thr Gln Gly Thr Phe Gly Ala Gly Arg Leu Lys Glu

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Gly Gly Val Asp Ser Cys Gln Gly Asp Ser Gly Gly Pro Leu Val Cys
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Met Ser Lys Thr Lys Asn Gly Ile Thr Cys Gln Lys Trp Ser Ser Thr
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Ser Pro His Arg Pro Arg Phe Ser Pro Ala Thr His Pro Ser Glu Gly
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Leu Glu Glu Asn Tyr Cys Arg Asn Pro Asp Asn Asp Pro Gln Gly Pro
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Trp Cys Tyr Thr Thr Asp Pro Glu Lys Arg Tyr Asp Tyr Cys Asp Ile
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Leu Glu Cys Glu Glu Cys Met His Cys Ser Gly Glu Asn Tyr Asp
                 85
ggc aaa att tcc aag acc atg tct gga ctg gaa tgc cag gcc tgg gac
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Gly Lys Ile Ser Lys Thr Met Ser Gly Leu Glu Cys Gln Ala Trp Asp
            100
                                105
tot cag ago coa cac got cat gga tac att cot too aaa ttt coa aac
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Ser Gln Ser Pro His Ala His Gly Tyr Ile Pro Ser Lys Phe Pro Asn
        115
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	tgg Trp															480
	ccc Pro														_	528
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	tcc Ser															624
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	agc Ser															768
	cca Pro	-		_	_		_	-			_					816
Thr	cct Pro	Val 275	Val	Gln	Asp	Cys	Tyr 280	His	Gly	Asp	Gly	Gln 285	Ser	Tyr	Arg	864
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ctg aaa aaa tgc tca gga aca gaa gcg Leu Lys Lys Cys Ser Gly Thr Glu Ala 355

325

1083

<210> 6 <211> 361 <212> PRT <213> Homo sapiens

<400> 6

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Pro Trp Cys Phe Thr Thr Asp Pro Ser Val Arg Trp Glu Tyr Cys Asn 340 345 Leu Lys Lys Cys Ser Gly Thr Glu Ala <210> 7 <211> 1086 <212> DNA <213> Mus musculus <220> <221> CDS <222> (1)...(1086) <400> 7 gtg tat ctg tca gaa tgt aag acc ggc atc ggc aac ggc tac aga gga 48 Val Tyr Leu Ser Glu Cys Lys Thr Gly Ile Gly Asn Gly Tyr Arg Gly 10 acc atg tcc agg aca aag agt ggt gtt gcc tgt caa aag tgg ggt gcc 96 Thr Met Ser Arg Thr Lys Ser Gly Val Ala Cys Gln Lys Trp Gly Ala 20 25 acg ttc ccc cac gta ccc aac tac tct ccc agt aca cat ccc aat gag 144 Thr Phe Pro His Val Pro Asn Tyr Ser Pro Ser Thr His Pro Asn Glu 40 gga cta gaa gag aac tac tgt agg aac cca gac aat gat gaa caa ggg 192 Gly Leu Glu Glu Asn Tyr Cys Arg Asn Pro Asp Asn Asp Glu Gln Gly 55 cct tgg tgc tac act aca gat ccg gac aag aga tat gac tac tgc aac 240 Pro Trp Cys Tyr Thr Thr Asp Pro Asp Lys Arg Tyr Asp Tyr Cys Asn att cct gaa tgt gaa gag gaa tgc atg tac tgc agt gga gaa aag tat 288 Ile Pro Glu Cys Glu Glu Glu Cys Met Tyr Cys Ser Gly Glu Lys Tyr gag ggc aaa atc tcc aag acc atg tct gga ctt gac tgc cag gcc tgg 336 Glu Gly Lys Ile Ser Lys Thr Met Ser Gly Leu Asp Cys Gln Ala Trp gat tot cag age cca cat get cat gga tac atc cct gcc aaa ttt cca 384 Asp Ser Gln Ser Pro His Ala His Gly Tyr Ile Pro Ala Lys Phe Pro age aag aac etg aag atg aat tat tge cae aac eet gae ggg gag eea 432 Ser Lys Asn Leu Lys Met Asn Tyr Cys His Asn Pro Asp Gly Glu Pro agg ccc tgg tgc ttc aca aca gac ccc acc aaa cgc tgg gaa tac tgt 480 Arg Pro Trp Cys Phe Thr Thr Asp Pro Thr Lys Arg Trp Glu Tyr Cys 150 155

gac Asp	atc Ile	ccc Pro	cgc Arg	tgc Cys 165	aca Thr	aca Thr	ccc Pro	ccg Pro	ccc Pro 170	cca Pro	ccc Pro	agc Ser	cca Pro	acc Thr 175	tac Tyr	528	}
caa Gln	tgt Cys	ctg Leu	aaa Lys 180	gga Gly	aga Arg	ggt Gly	gaa Glu	aat Asn 185	tac Tyr	cga Arg	gly ggg	acc Thr	gtg Val 190	tct Ser	gtc Val	57€	;
acc Thr	gtg Val	tct Ser 195	GJA aaa	aaa Lys	acc Thr	tgt Cys	cag Gln 200	cgc Arg	tgg Trp	agt Ser	gag Glu	caa Gln 205	acc Thr	cct Pro	cat His	624	ŧ
agg Arg	cac His 210	aac Asn	agg Arg	aca Thr	cca Pro	gaa Glu 215	aat Asn	ttc Phe	ccc Pro	tgc Cys	aaa Lys 220	aat Asn	ctg Leu	gaa Glu	gag Glu	672	2
aac Asn 225	tac Tyr	tgc Cys	cgg Arg	aac Asn	cca Pro 230	gat Asp	gga Gly	gaa Glu	act Thr	gct Ala 235	ccc Pro	tgg Trp	tgc Cys	tat Tyr	acc Thr 240	720)
act Thr	gac Asp	agc Ser	cag Gln	ctg Leu 245	agg Arg	tgg Trp	gag Glu	tac Tyr	tgt Cys 250	gag Glu	att Ile	cca Pro	tcc Ser	tgc Cys 255	gag Glu	768	3
	tca Ser															816	5
	aca Thr															864	4
	ggt Gly 290															91:	2
gca Ala 305	gct Ala	atg Met	ttt Phe	cca Pro	cac His 310	agg Arg	cat His	tcg Ser	aag Lys	acc Thr 315	cca Pro	gag Glu	aac Asn	ttc Phe	cca Pro 320	96	0
gat Asp	gct Ala	ggc	ttg Leu	gag Glu 325	atg Met	aac Asn	tac Tyr	Cys	agg Arg 330	Asn	ccg Pro	gat Asp	ggt Gly	gac Asp 335	Lys	100	8
ggc	cct Pro	tgg Trp	tgc Cys 340	Tyr	acc Thr	act Thr	gac Asp	ccg Pro 345	Ser	gtc Val	agg Arg	tgg Trp	gaa Glu 350	Tyr	tgc Cys	105	6
	ctg Leu		Arg					Gly								108	6

<210> 8 <211> 362

<212> PRT <213> Mus musculus

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<210> 9 <211> 552 <212> DNA <213> Mus musculus

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acc Thr	ccc Pro	ctg Leu	tct Ser 20	gga Gly	ggc Gly	atg Met	cgt Arg	ggt Gly 25	atc Ile	cgt Arg	gga Gly	gca Ala	gat Asp 30	ttc Phe	cag Gln	96
tgc Cys	ttc Phe	cag Gln 35	caa Gln	gcc Ala	cga Arg	gcc Ala	gtg Val 40	Gly 999	ctg Leu	tcg Ser	ggc Gly	acc Thr 45	ttc Phe	cgg Arg	gct Ala	144
ttc Phe	ctg Leu 50	tcc Ser	tct Ser	agg Arg	ctg Leu	cag Gln 55	gat Asp	ctc Leu	tat Tyr	agc Ser	atc Ile 60	gtg Val	cgc Arg	cgt Arg	gct Ala	192
gac Asp 65	cgg Arg	Gly 999	tct Ser	gtg Val	ccc Pro 70	atc Ile	gtc Val	aac Asn	ctg Leu	aag Lys 75	gac Asp	gag Glu	gtg Val	cta Leu	tct Ser 80	240
ccc Pro	agc Ser	tgg Trp	gac Asp	tcc Ser 85	ctg Leu	ttt Phe	tct Ser	ggc Gly	tcc Ser	cag Gln	ggt Gly	caa Gln	gtg Val	caa Gln 95	ccc Pro	288
Gly 999	gcc Ala	cgc Arg	atc Ile 100	ttt Phe	tct Ser	ttt Phe	gac Asp	ggc Gly 105	aga Arg	gat Asp	gtc Val	ctg Leu	aga Arg 110	cac His	cca Pro	336
gcc Ala	tgg Trp	ccg Pro 115	Gln	aag Lys	agc Ser	gta Val	tgg Trp 120	His	ggc Gly	tcg Ser	gac Asp	ccc Pro 125	Ser	gly aaa	cgg Arg	384
agg Arg	ctg Leu 130	Met	gag Glu	agt Ser	tac Tyr	tgt Cys 135	gag Glu	aca Thr	tgg Trp	cga Arg	act Thr 140	Glu	act Thr	act Thr	gly	432
gct Ala 145	aca Thr	ggt Gly	cag Gln	gcc Ala	tcc Ser 150	Ser	ctg	ctg Leu	tca Ser	ggc Gly 155	/ Arg	cto Lev	ctg Leu	gaa Glu	cag Gln 160	480
aaa Lys	gct Ala	gcg Ala	g ago a Ser	tgo Cys	His	aac Asn	ago Ser	tac Tyr	ato 110	· Val	ctg L Leu	tgo Cys	att	gaç Glu 175	ı Asn	528
	ttc Phe			Sei												552

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            20
Cys Phe Gln Gln Ala Arg Ala Val Gly Leu Ser Gly Thr Phe Arg Ala
Phe Leu Ser Ser Arg Leu Gln Asp Leu Tyr Ser Ile Val Arg Arg Ala
                        55
                                            60
Asp Arg Gly Ser Val Pro Ile Val Asn Leu Lys Asp Glu Val Leu Ser
                                        75
                    70
Pro Ser Trp Asp Ser Leu Phe Ser Gly Ser Gln Gly Gln Val Gln Pro
                                    90
                85
Gly Ala Arg Ile Phe Ser Phe Asp Gly Arg Asp Val Leu Arg His Pro
                                                    110
                                105
            100
Ala Trp Pro Gln Lys Ser Val Trp His Gly Ser Asp Pro Ser Gly Arg
                                                125
                            120
Arg Leu Met Glu Ser Tyr Cys Glu Thr Trp Arg Thr Glu Thr Thr Gly
                                            140
                        135
Ala Thr Gly Gln Ala Ser Ser Leu Leu Ser Gly Arg Leu Leu Glu Gln
                                    155
Lys Ala Ala Ser Cys His Asn Ser Tyr Ile Val Leu Cys Ile Glu Asn
                                   170
Ser Phe Met Thr Ser Phe Ser Lys .
            180
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Met Asp His Lys Glu Val Ile Leu Leu Phe Leu Leu Leu Lys Pro
                                      10
 1
gga caa ggg gac tcg ctg gat ggc tac ata agc aca caa ggg gct tca
                                                                       96
Gly Gln Gly Asp Ser Leu Asp Gly Tyr Ile Ser Thr Gln Gly Ala Ser
                                  25
 ctg ttc agt ctc acc aag aag cag ctc gca gca gga ggt gtc tcg gac
                                                                      144
 Leu Phe Ser Leu Thr Lys Lys Gln Leu Ala Ala Gly Gly Val Ser Asp
                              4 C
      . 35
 tgt ttg gcc aaa tgt gaa ggg gaa aca gac ttt gtc tgc agg tca ttc
                                                                      192
 Cys Leu Ala Lys Cys Glu Gly Glu Thr Asp Phe Val Cys Arg Ser Phe
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													gag Glu			240
													ttc Phe			288
													ggc Gl <u>y</u> 110			336
gga Gly	acc Thr	atg Met 115	tcc Ser	agg Arg	aca Thr	aag Lys	agt Ser 120	ggt Gly	gtt Val	gcc Ala	tgt Cys	caa Gln 125	aag Lys	tgg Trp	ggt Gly	384
													cat His			432
													gat Asp			480
													gac Asp			528
aac Asn	att Ile	cct Pro	gaa Glu 180	tgt Cys	gaa Glu	gag Glu	gaa Glu	tgc Cys 185	atg Met	tac Tyr	tgc Cys	agt Ser	gga Gly 190	gaa Glu	aag Lys	576
Tyr	Glu	Gly 195	Lys	Ile	Ser	Lys	Thr 200	Met	Ser	Gly	Leu	Asp 205	tgc Cys	Gln	Ala	624
Trp	Asp 210	Ser	Gln	Ser	Pro	His 215	Ala	His	Gly	Tyr	11e 220	Pro	gcc Ala	Lys	Phe	672
Pro 225	Ser	Lys	Asn	Leu	Lys 230	Met	Asn	Tyr	Cys	His 235	Asn	Pro	gac Asp	Gly	Glu 240	720
Pro	Arg	Pro	Trp	Cys 245	Phe	Thr	Thr	Asp	Pro 250	Thr	Lys	Arg	tgg Trp	Glu 255	Tyr	768
Cys	Asp	Ile	Pro 260	Arg	Cys	Thr	Thr	Pro 265	Pro	Pro	Pro	Pro	agc Ser 270	Pro	Thr	816
tac Tyr	caa Gln	tgt Cys	ctg Leu	aaa Lys	gga Gly	aga Arg	ggt Gly	gaa Glu	aat Asn	tac Tyr	cga Arg	Gly	acc Thr	gtg Val	tct Ser	864

	275					280					285					
gtc ac Val Th	r Val															912
cat ag His Ar 305																960
gag aa Glu As															1	800
acc ac															1	056
gag to Glu Se															1	104
gag ca Glu Gl 37	n Thr														1	152
tat cg Tyr Ar 385	g Gly	aca Thr	tcg Ser	ser 390	act Thr	acc Thr	atc Ile	aca Thr	999 Gly 395	aag Lys	aag Lys	tgc Cys	cag Gln	tcc Ser 400	1	200
tgg gö Trp Al	a gct a Ala	atg Met	ttt Phe 405	cca Pro	cac His	agg Arg	càt His	tcg Ser 410	aag Lys	acc Thr	cca Pro	gag Glu	aac Asn 415	ttc Phe	1	248
cca ga Pro As															1	296
aag gg Lys Gl	gc cct y Pro 435	tgg Trp	tgc Cys	tac Tyr	acc Thr	act Thr 440	gac Asp	ccg Pro	agc Ser	gtc Val	agg Arg 445	tgg Trp	gaa Glu	tac Tyr	1	344
tgc aa Cys As 45	n Leu														1	.392
gac ga Asp As 465	_	-			taa *	С									1	.414

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Leu Phe Ser Leu Thr Lys Lys Gln Leu Ala Ala Gly Gly Val Ser Asp
Cys Leu Ala Lys Cys Glu Gly Glu Thr Asp Phe Val Cys Arg Ser Phe
Gln Tyr His Ser Lys Glu Gln Gln Cys Val Ile Met Ala Glu Asn Ser
                   70
Lys Thr Ser Ser Ile Ile Arg Met Arg Asp Val Ile Leu Phe Glu Lys
                                   90
Arq Val Tyr Leu Ser Glu Cys Lys Thr Gly Ile Gly Asn Gly Tyr Arg
                               105
Gly Thr Met Ser Arg Thr Lys Ser Gly Val Ala Cys Gln Lys Trp Gly
                           120
Ala Thr Phe Pro His Val Pro Asn Tyr Ser Pro Ser Thr His Pro Asn
                       135
                                           140
Glu Gly Leu Glu Glu Asn Tyr Cys Arg Asn Pro Asp Asn Asp Glu Gln
                   150
                                       155
Gly Pro Trp Cys Tyr Thr Thr Asp Pro Asp Lys Arg Tyr Asp Tyr Cys
                                  170
               165
Asn Ile Pro Glu Cys Glu Glu Cys Met Tyr Cys Ser Gly Glu Lys
                               185
Tyr Glu Gly Lys Ile Ser Lys Thr Met Ser Gly Leu Asp Cys Gln Ala
                           200
Trp Asp Ser Gln Ser Pro His Ala His Gly Tyr Ile Pro Ala Lys Phe
                      215
                                            220
Pro Ser Lys Asn Leu Lys Met Asn Tyr Cys His Asn Pro Asp Gly Glu
                   230
                                       235
Pro Arg Pro Trp Cys Phe Thr Thr Asp Pro Thr Lys Arg Trp Glu Tyr
                                   250
Cys Asp Ile Pro Arg Cys Thr Thr Pro Pro Pro Pro Pro Ser Pro Thr
                               265
Tyr Gln Cys Leu Lys Gly Arg Gly Glu Asn Tyr Arg Gly Thr Val Ser
                           280
Val Thr Val Ser Gly Lys Thr Cys Gln Arg Trp Ser Glu Gln Thr Pro
                                           300
                        295
His Arg His Asn Arg Thr Pro Glu Asn Phe Pro Cys Lys Asn Leu Glu
                   310
                                        315
Glu Asn Tyr Cys Arg Asn Pro Asp Gly Glu Thr Ala Pro Trp Cys Tyr
                                   330
               325
Thr Thr Asp Ser Gln Leu Arg Trp Glu Tyr Cys Glu Ile Pro Ser Cys
                               345
           340
Glu Ser Ser Ala Ser Pro Asp Gln Ser Asp Ser Ser Val Pro Pro Glu
                           360
Glu Gln Thr Pro Val Val Gln Glu Cys Tyr Gln Ser Asp Gly Gln Ser
                       375
Tyr Arg Gly Thr Ser Ser Thr Thr Ile Thr Gly Lys Lys Cys Gln Ser
                   390
                                        395
Trp Ala Ala Met Phe Pro His Arg His Ser Lys Thr Pro Glu Asn Phe
                                   410
Pro Asp Ala Gly Leu Glu Met Asn Tyr Cys Arg Asn Pro Asp Gly Asp
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145			150			155			160	
	cgg Arg									528
	Gly 999									576
	cag Gln									624
	aat Asn 210						С			661

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<211> 218

<212> PRT

<213> Mus musculus

<400> 14

Met Asp His Lys Glu Val Ile Leu Leu Phe Leu Leu Leu Leu Lys Pro Gly Gln Gly Asp Ser Leu Asp Leu Asp Tyr Lys Asp Asp Asp Asp Lys Leu Ala His Thr His Gln Asp Phe Gln Pro Val Leu His Leu Val Ala Leu Asn Thr Pro Leu Ser Gly Gly Met Arg Gly Ile Arg Gly Ala Asp Phe Gln Cys Phe Gln Gln Ala Arg Ala Val Gly Leu Ser Gly Thr Phe 70 Arg Ala Phe Leu Ser Ser Arg Leu Gln Asp Leu Tyr Ser Ile Val Arg Arg Ala Asp Arg Gly Ser Val Pro Ile Val Asn Leu Lys Asp Glu Val 105 Leu Ser Pro Ser Trp Asp Ser Leu Phe Ser Gly Ser Gln Gly Gln Val 120 Gln Pro Gly Ala Arg Ile Phe Ser Phe Asp Gly Arg Asp Val Leu Arg 135 His Pro Ala Trp Pro Gln Lys Ser Val Trp His Gly Ser Asp Pro Ser 155 150 Gly Arg Arg Leu Met Glu Ser Tyr Cys Glu Thr Trp Arg Thr Glu Thr 170 165 Thr Gly Ala Thr Gly Gln Ala Ser Ser Leu Leu Ser Gly Arg Leu Leu 185 190 Glu Gln Lys Ala Ala Ser Cys His Asn Ser Tyr Ile Val Leu Cys Ile 200 Glu Asn Ser Phe Met Thr Ser Phe Ser Lys 210 215

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		180					185					190			
tat gag Tyr Glu															624
tgg gat Trp Asp 210	Ser														672
cca agc Pro Ser 225															720
cca agg Pro Arg															768
tgt gac Cys Asp															816
tac caa Tyr Gln															864
gtc acc Val Thr 290	Val														912
cat agg His Arg 305															960
gag aac Glu Asr															1008
acc act															1056
gag tco Glu Ser															1104
gag caa Glu Glr 370	Thr														1152
gat ctt Asp Lei 385	gac Asp	tac Tyr	aag Lys	gac Asp 390	gac Asp	gat Asp	gac Asp	aag Lys	ctt Leu 395	gct Ala	cat His	act Thr	cat His	cag Gln 400	1200
gac ttt	cag	cca	gtg	ctc	cac	ctg	gtg	gca	ctg	aac	acc	ccc	ctg	tct	1248

Asp	Phe	Gln	Pro	Val 405	Leu	His	Leu	Val	Ala 410	Leu	Asn	Thr	Pro	Leu 415	Ser		
		_	_			_		_			cag Gln	_		_			1296
_	_										gct Ala						1344
	_	_	_			_			-	_	gct Ala 460	-					1392
											tct Ser						1440
	_					_					ccc Pro		_	_			1488
			-		_		_	-	_		cca Pro	_		-	_	•	1536
_	_	-				_	_		_		cgg Arg		_	_			1584
_		_				_		-			999 Gly 540	_			_		1632
_			_						_	_	cag Gln		_		_		1680
_			_			-	_	_			aat Asn	_		_			1728
	ttc Phe			taa *	taa *	С	٠										1747

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<211> 580 <212> PRT

<213> Mus musculus

<400> 16

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Leu Phe Ser Leu Thr Lys Lys Gln Leu Ala Ala Gly Gly Val Ser Asp
Cys Leu Ala Lys Cys Glu Gly Glu Thr Asp Phe Val Cys Arg Ser Phe
Gln Tyr His Ser Lys Glu Gln Gln Cys Val Ile Met Ala Glu Asn Ser
                                       75
Lys Thr Ser Ser Ile Ile Arg Met Arg Asp Val Ile Leu Phe Glu Lys
                                   90
Arg Val Tyr Leu Ser Glu Cys Lys Thr Gly Ile Gly Asn Gly Tyr Arg
                               105
Gly Thr Met Ser Arg Thr Lys Ser Gly Val Ala Cys Gln Lys Trp Gly
                                               125
                           120
Ala Thr Phe Pro His Val Pro Asn Tyr Ser Pro Ser Thr His Pro Asn
                                           140
                       135
Glu Gly Leu Glu Glu Asn Tyr Cys Arg Asn Pro Asp Asn Asp Glu Gln
                                       155
                   150
Gly Pro Trp Cys Tyr Thr Thr Asp Pro Asp Lys Arg Tyr Asp Tyr Cys
                                   170
Asn Ile Pro Glu Cys Glu Glu Glu Cys Met Tyr Cys Ser Gly Glu Lys
           180
                                185
Tyr Glu Gly Lys Ile Ser Lys Thr Met Ser Gly Leu Asp Cys Gln Ala
                           200
Trp Asp Ser Gln Ser Pro His Ala His Gly Tyr Ile Pro Ala Lys Phe
                       215
Pro Ser Lys Asn Leu Lys Met Asn Tyr Cys His Asn Pro Asp Gly Glu
                                       235
                   230
Pro Arg Pro Trp Cys Phe Thr Thr Asp Pro Thr Lys Arg Trp Glu Tyr
                245
                                    250
Cys Asp Ile Pro Arg Cys Thr Thr Pro Pro Pro Pro Pro Ser Pro Thr
                                265
Tyr Gln Cys Leu Lys Gly Arg Gly Glu Asn Tyr Arg Gly Thr Val Ser
                                                285
                            280
Val Thr Val Ser Gly Lys Thr Cys Gln Arg Trp Ser Glu Gln Thr Pro
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PCT/US98/24950 WO 99/26480

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INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/24950

IPC(6)	IPC(6) :A01N 63/00, 43/04; C12N 15/00; C07H 21/02									
	: 424/93.1; 435/320.1; 514/44; 536/23.1 to International Patent Classification (IPC) or to both	national classification and IPC								
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Minimum d	locumentation searched (classification system follower	ed by classification symbols)								
U.S. :	424/93.1; 435/320.1; 514/44; 536/23.1									
Documenta	tion searched other than minimum documentation to th	e extent that such documents are included	in the fields searched							
Electronic d	lata base consulted during the international search (n	ame of data base and, where practicable,	search terms used)							
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C. DOCUMENTS CONSIDERED TO BE RELEVANT										
Category*	Citation of document, with indication, where ap	ppropriate, of the relevant passages	Relevant to claim No.							
Y,P	US 5,792,845 A (O'REILLY et al.) 11 August 1998 (11.08.98), 1-30, 33 col. 4, lines 32-68, col. 5, lines 1-2, 51-68, col. 6, lines 1-8.									
Y	WO 97/23500 A1 (THE CHILDREN'S MEDICAL CENTER 4 CORPORATION) 03 July 1997 (03.07.97), page 41, lines 3-33, page 42, lines 1-27.									
X,P	WO 98/49321 A2 (RHONE-POULE 1998 (05.11.98), page 44, 6-11, 25-33		1, 5, 18, 20, 31							
Y,P	1990 (03:11:50), page 44, 0-11, 23-33	, page 43, fines 12-13, 23-33.	2-4, 6-17, 19, 21- 30, 32, 33							
X Furth	er documents are listed in the continuation of Box C	C. See patent family annex.								
A doc	ecial categories of cited documents: cument defining the general state of the art which is not considered be of particular relevance	"T" later document published after the inte date and not in conflict with the appl the principle or theory underlying the	ication but cited to understand							
"L" doc	lier document published on or after the international filing data cument which may throw doubts on priority claim(s) or which is ad to establish the publication data of another citation or other	"X" document of particular relevance; the considered novel or cannot be conside when the document is taken alone								
O doc	eisl reason (as specified) cument referring to an oral disclosure, use, exhibition or other ans	"Y" document of particular relevance; the considered to involve an inventive combined with one or more other such being obvious to a person skilled in t	step when the document is a documents, such combination							
P doc	nument published prior to the international filing date but later than priority date claimed	*&* document member of the same patent	family							
1.4	actual completion of the international search JARY 1999	Date of mailing of the international sea 0.8 MAR 19	•							
Commission Box PCT Washington	nailing address of the ISA/US ner of Patents and Trademarks D.C. 20231	Authorized office SHIN-LIN CHEN	7 1100/0							
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INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/24950

	PC1/US98/24	
C (Continu	ation). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WO 97/15666 A (THE CHILDREN'S MEDICAL CENTER CORPORATION) 01 May 1997 (01.05.97), page 20, lines 16-35, page 21, page 22, lines 1-16. page 59, lines 5-35, page 60, page 61, 1-6.	1-30, 33
Y	TANAKA, T. et al. Retroviral and adenoviral mediated transduction of angiostatin cDNA inhibits angiogenesis and tumor growth. Proceedings of the American Association for Cancer Research. March 1997 (03.97). Vol 38. page 264.	1-33
Y	WO 96/35774 A2 (THE CHILDREN'S MEDICAL CENTER CORPORATION) 14 November 1996 (14.11.96), page lines 33-36, pages 22-25, page 26, lines 1-33. pages 144-148.	1-30, 33
Y	WO 97/41824 A2(ABBOTT LABORATORIES) 13 November 1997 (13.11.97), page 5, lines 13-38, page 6, 1-18, page 60, lines 15-38, pages 61-62, page 63, lines 1-33.	1-30, 33
Y	WO 95/29242 A1 (THE CHILDREN; S MEDICAL CENTER CORPORATION) 02 November 1995 (02.11.95), page 21, lines 19-35, pages 22-27, page 28, lines 1-4. page 87, lines 4-35, page 88, page 89, lines 1-14.	1-30, 33
Y	O'REILLY, et al. Angiostatin induces and sustains dormancy of human primary tumors in mice. Nature Medicine, June 1996 (06.96), Vol. 2, No. 6, pages 689-692, especially pages 689-690.	1-30, 33
1	O'REILLY, et al. Endostatin: An Endogenous Inhibitor of Angiogenesis and Tumor Growth. Cell, 24 January 1997 (24.01.97), Vol. 88, pages 277-285, especially pages 279-280, 282.	1-30, 33

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INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/24950

B. FIELDS SEARCHED Electronic data bases consulted (N	lame of data base and where prac	cticable terms used):	
APS, STN, WPIDS, MEDLINE, of search terms: angiostatin, plasmino diabet?(p)retinopathy, plasmid, vir.	ogen, endostatin, colagen(w) XVII	II, inhibit?(5a)tumor(5a)growth, tumor(5a)regress'	?.
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